

7,8-Dihydro Analogs of Ecdysteroids*

V. N. Odínokov^a, S. R. Afon'kina^a, R. V. Shafikov^a, R. G. Savchenko^a, I. V. Galyautdinov^a,
L. M. Khalilov^a, and A. S. Shashkov^b

^a Institute of Petroleum Chemistry and Catalysis, Russian Academy of Sciences,
pr. Oktyabrya 141, Ufa, 450075 Bashkortostan, Russia
e-mail: ink@anrb.ru

^b Zelinskii Institute of Organic Chemistry, Russian Academy of Sciences, Moscow, Russia

Received August 2, 2006

Abstract—Reactions of 20-hydroxyecdysone, its diacetonide, and 24,25(25,26)-anhydro derivative with lithium tetrahydridoaluminate gave the corresponding 6 α - and 6 β -epimeric alcohols and 7,8-dihydro analogs.

DOI: 10.1134/S107042800706005X

Selective reduction of the double Δ^7 -bonds in ecdysteroids with formation of 7,8-dihydro analogs opens a way to new biologically active structures. Catalytic hydrogenation of ecdysteroids does not produce the desired products because of numerous side reactions [2–4]. Suksamrarn et al. [5] recently reported on stereoselective catalytic hydrogenation of Δ^7 -6-oxo steroids in the presence of sodium nitrite; however, we failed to obtain 7,8-dihydro analogs from 20-hydroxyecdysone and its derivatives according to the procedure described in [5]. Our attempts to hydrogenate the Δ^7 -bond in 20-hydroxyecdysone and its derivatives via reaction with alkali metals in liquid ammonia [6, 7] (which is used for selective reduction of double bond in conjugated ketones of the steroid series) were also unsuccessful.

While studying the reactions of 20-hydroxyecdysone (**I**), its 2,3:20,22-diacetonide **II** and 24,25-(25,26)-anhydro-20-hydroxyecdysone diacetonide **III** (a 3:2 mixture of Δ^{24} and Δ^{25} derivatives) with lithium tetrahydridoaluminate we revealed formation of the corresponding 7,8-dihydro analogs **IV–VI** (Scheme 1). In the reaction of compound **I** with LiAlH_4 at a molar ratio of 1:7 (a suspension in diethyl ether) we isolated 21% of 7,8-dihydro analog **IV** which was identical in the IR and NMR (^1H and ^{13}C) spectra to a sample described previously [5]. The reactions of diacetonides **II** and **III** with 3 equiv of LiAlH_4 gave the corresponding 7,8-dihydro analogs **V** and **VI** in 35 and 30% yield, respectively. In addition, we isolated mixtures (~1:2)

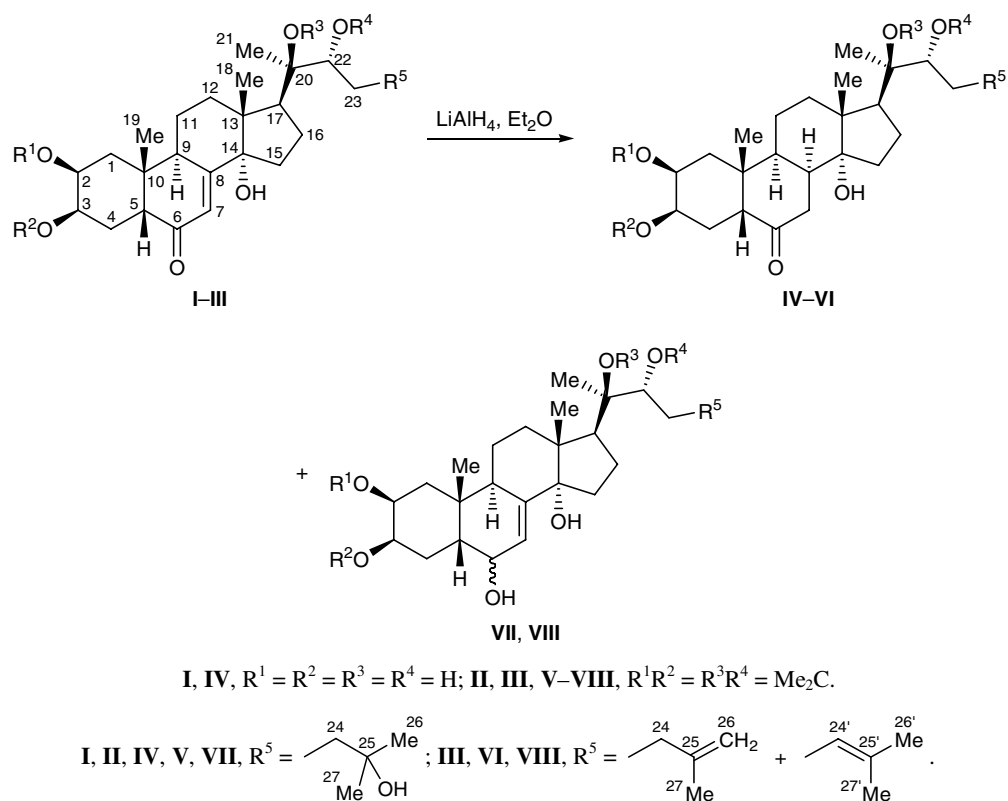
of previously described [8] 6 α /6 β -epimeric alcohols **VII** and **VIII**, whose yields were 60 and 65%, respectively. When the reaction with diacetonide **II** was carried out by adding LiAlH_4 to a solution of **II** (i.e., the order of mixing the reactants was changed), ~10% of compound **IX** was formed together with **V** and **VII**. The reaction with 5 equiv of LiAlH_4 gave rise to a mixture of compounds **V**, **VII**, and **X** (Scheme 2).

It could be presumed that 7,14-diene **IX** is formed via dehydration of alcohol **VII** and that $\Delta^{14(15)}$ -alcohol **X** is the product of reduction and dehydration of 7,8-dihydro derivative **V**. However, compounds **IX** and **X** were not obtained in the reactions of alcohols **VII** and **V** with LiAlH_4 . In these cases, alcohol **VII** was recovered from the reaction mixture, while the reaction of ketone **V** with LiAlH_4 afforded the corresponding alcohol **XI** which failed to react with LiAlH_4 . Therefore, we concluded that compounds **IX** and **X** were formed from a common precursor.

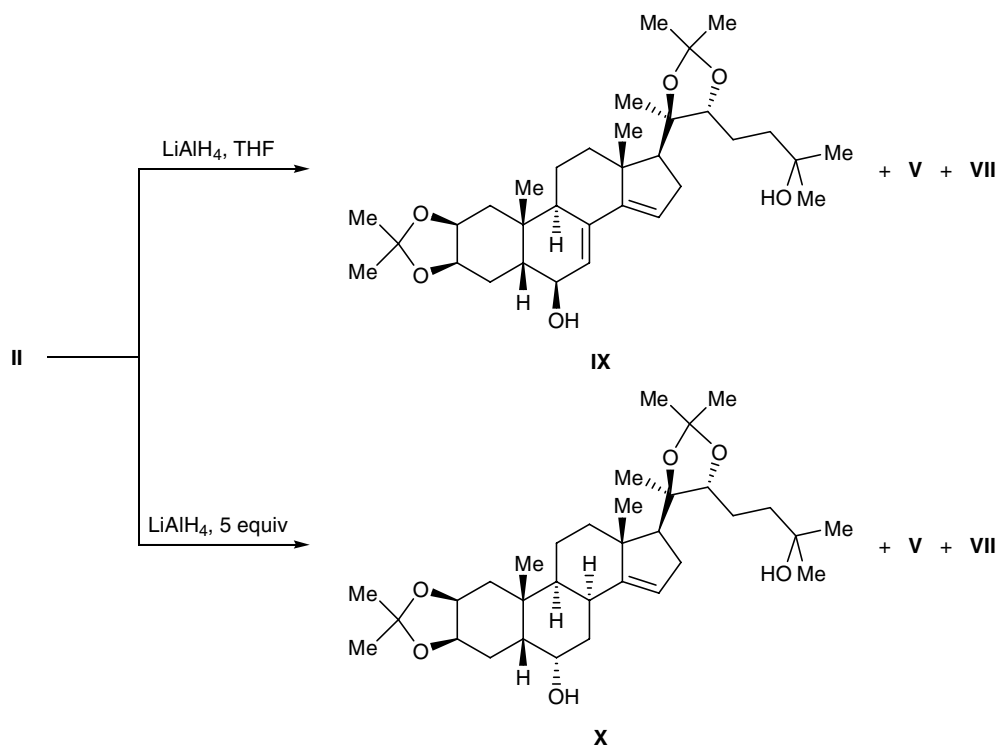
The ^{13}C NMR spectra of saturated ketones **IV–VI** displayed characteristic differences from the spectra of initial conjugated enones **I–III**. The C^7 and C^8 signals appeared in a considerably stronger field, while the C^6 signal was displaced downfield ($\Delta\delta_{\text{C}} \sim 9$ ppm) due to the lack of conjugation between the carbonyl group and double bond. Signals in the ^1H and ^{13}C NMR spectra of ketone **V** and alcohol **XI** were assigned using homo- and heteronuclear correlation techniques (COSY, HSQC, HMBC, TOCSY, ROESY). ROESY experiments showed nuclear Overhauser effect between protons of the β -oriented C^{19}H_3 group and 5-H

* For preliminary communication, see [1].

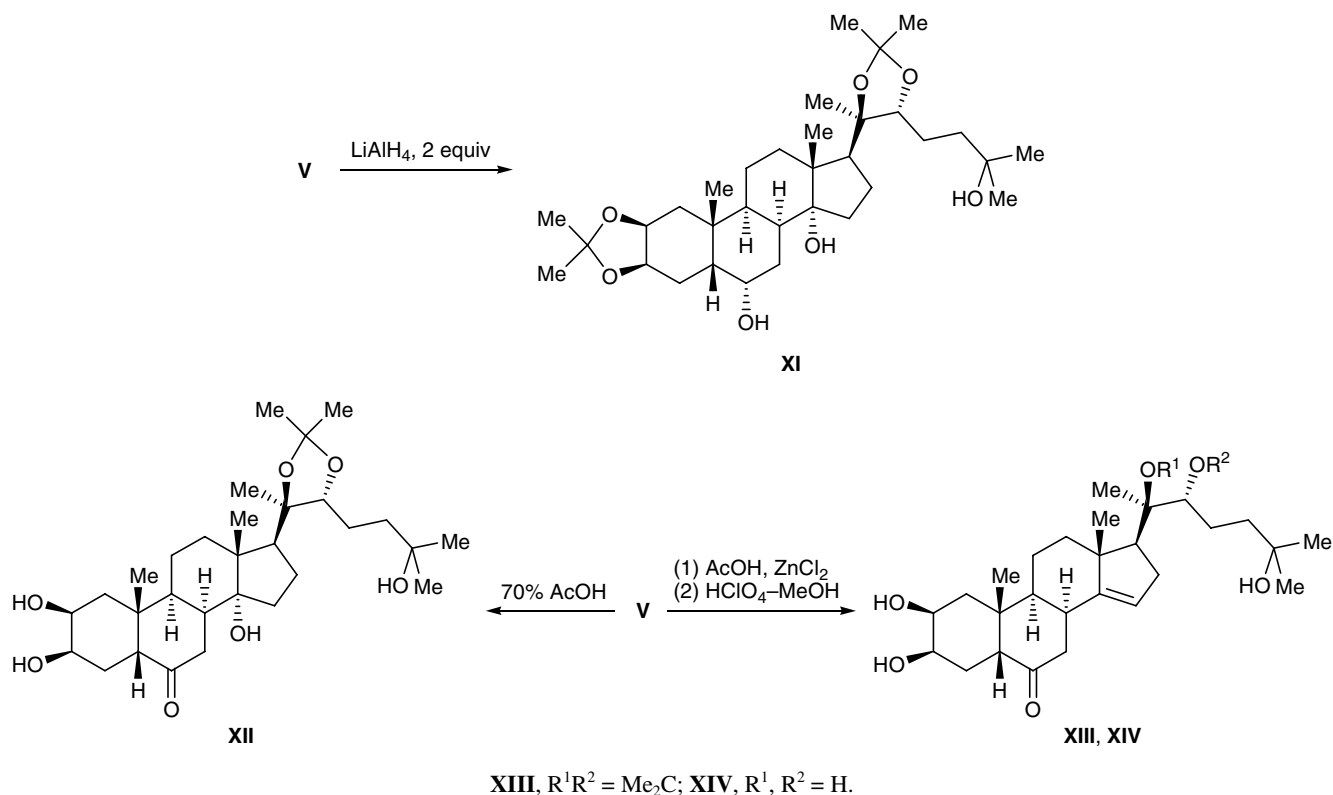
Scheme 1.



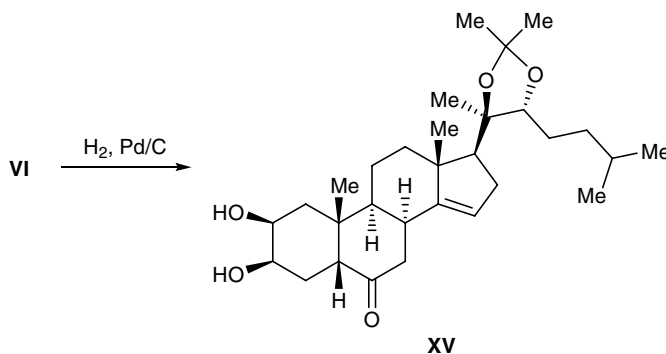
Scheme 2.



Scheme 3.



Scheme 4.

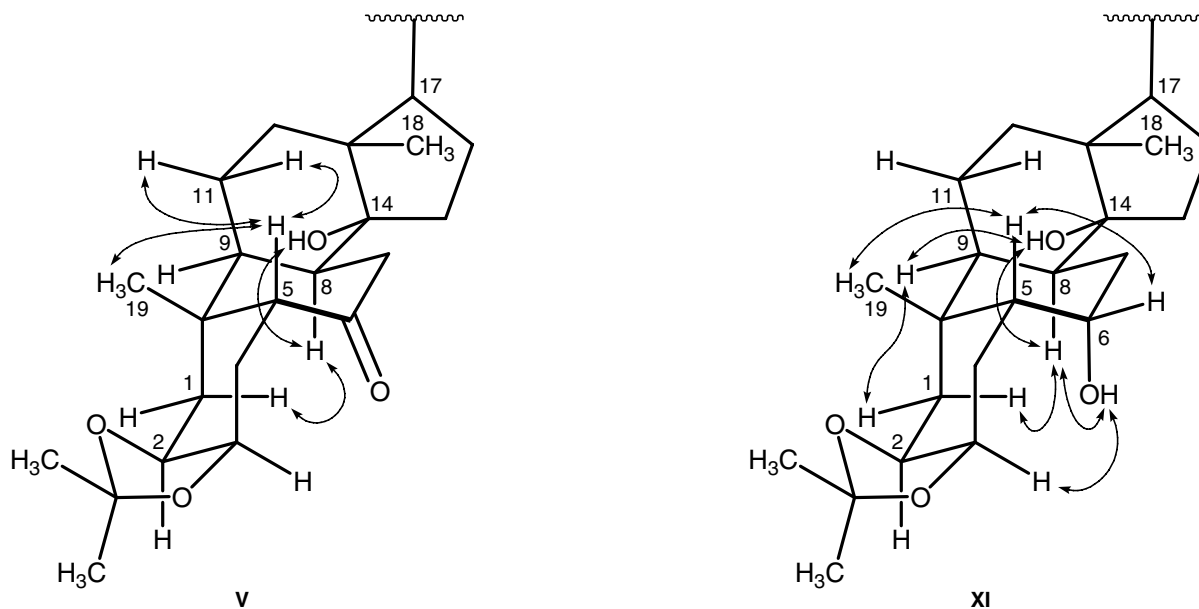


in molecule **V**, indicating *cis*-junction of the A and B rings, i.e., β -orientation of 5-H. The existence of NOE between 8-H and α -14-OH (ROESY) confirmed α -orientation of the former (see figure).

These data show that hydride reduction of the Δ^7 -bond in diacetonide **II** is not accompanied by epimerization at C⁵ and that the attack by hydride ion on the double-bonded C⁸ atom occurs at the α -side. Unlike α,β -unsaturated ketone **II** [8], no epimerization at C⁵ was observed in the hydride reduction of the ketone group in unsaturated ketone **V**. The reduction of ketone **V** with lithium tetrahydridoaluminate is stereospecific, and the product is 6α -alcohol **XI** (Scheme 3),

while the reduction of the 6-oxo group in compound **II** under the same conditions gives a 1:2 mixture of 6α - and 6β -epimeric alcohols **VII** [8].

The structure of alcohol **XI** (see figure) was determined on the basis of NOE cross peaks observed in the ROESY spectra. β -Orientation of the 5-H proton follows from the existence of a correlation between that proton and protons of the C¹⁹H₃ group (HMBC). Couplings between the 14-OH proton, on the one hand, and 8-H and 9-H, on the other, suggests α -configuration of the newly formed chiral center at C⁸, and a strong cross peak between 5-H and 6-H unambiguously indicates β -orientation of the latter and hence



Cross couplings (NOEs) in the ROESY experiments for compounds **V** and **XI**.

formation of 6 α -epimeric alcohol **XI**. An additional support to the α -orientation of the 6-OH group is provided by the presence of a cross peak between the 6-OH proton and 8-H. Interaction between the 6 α -OH proton and 3-H was also observed; this means that the latter occupies axial position; correspondingly, the 2-H proton is oriented equatorially.

As follows from our data on the stereochemistry of hydride reduction, the known C⁵-epimerization of ecdysteroids under alkaline conditions [9, 10] accompanies the reduction of the 6-oxo group in Δ^7 -6-oxo ecdysteroids; as a result, 6-hydroxy derivatives of the 5 α -series are formed [8]. The hydride reduction of the Δ^7 -bond in Δ^7 -6-oxo ecdysteroids **I–III** gives rise to 7,8-dihydro derivatives **IV–VI** of the 5 β -series. Likewise, no epimerization at C⁵ is observed in the hydrogenation of the oxo group in saturated ketone **V**. We can conclude that 5 β -epimers of 7,8-dihydro-6-oxo and 7,8-dihydro-6-hydroxy ecdysteroid derivatives with *cis*-junction of the A and B rings are more stable.

The structure of 7,14-dien-6 β -ol **IX** is confirmed by comparison of its ¹H and ¹³C NMR spectra with those of the corresponding 6 α -epimer [8]. Signals from C⁵ and C⁶ in the ¹³C NMR spectrum of the former appear in a weaker field ($\Delta\delta_C = 1.8$ and 4.1 ppm, respectively, while the C⁷ signal is displaced upfield ($\Delta\delta_C = 1.0$ ppm; the C⁷ and C¹⁵ signals were misassigned in [8]). The 6-H signal in the ¹H NMR spectrum of 6 β -dienol **IX** is located in a stronger field ($\Delta\delta = 0.76$ ppm) relative to the corresponding signal of the 6 α -epimer (cf. the data for 6 α - and 6 β -epimeric alcohols **VII** [8]).

The presence of double C¹⁴=C¹⁵ bond in alcohol **X** induces appreciable changes in the ¹H and ¹³C NMR spectral patterns as compared to saturated alcohol **XI**. The ¹H NMR spectrum of **X** contains a signal at δ 5.35 ppm from the vinyl proton on C¹⁵, and the C¹⁴ and C¹⁵ *sp*²-hybridized carbon atoms give rise to downfield signals at δ_C 154.4 and 120.6 ppm, respectively, in the ¹³C NMR spectrum. The C⁸, C¹², C¹⁶, and C¹⁷ signals are displaced downfield ($\Delta\delta_C = 11.5$, 7.4, 7.0, and 9.7 ppm, respectively), while the C⁹ signal is located in a stronger field ($\Delta\delta_C = 11.3$ ppm). As follows from the Dreiding model, the C⁸, C¹⁶, and C¹⁷ atoms in molecule **X** lie in the C¹⁴=C¹⁵ bond plane and are therefore deshielded, while the C⁹ atom falls into the shielded area (out of the double bond plane) [11].

Hydrolysis of diacetone **V** in 70% acetic acid or AcOH/ZnCl₂ [12] resulted in deprotection of only hydroxy groups on C² and C³; the reaction in the presence of ZnCl₂ was accompanied by dehydration to generate $\Delta^{14(15)}$ -bond. As a result, 20,22-acetonides **XII** and **XIII** were obtained, respectively. Deprotection of all four hydroxy groups was achieved using perchloric acid in methanol [13]. In this case, the reaction was also accompanied by dehydration, and the product was enone **XIV** (Scheme 3).

The formation of $\Delta^{14(15)}$ bond also occurred in the catalytic hydrogenation of the side-chain double bond of Δ^{24} (Δ^{25})-alkenes **VI**. The reaction was accompanied by deprotection of the hydroxy groups on C² and C³ to give enone **XV** (Scheme 4). An analogous process was

described in [4]. Deprotection of the 2,3-dihydroxy moiety is readily identified by the ^{13}C NMR spectra of compounds **XII**, **XIII**, and **XV**, which contain only one acetal signal at δ_{C} 106.8 ppm. Deshielding effect of the $\Delta^{14(15)}$ -bond leads to downfield shift of the 8-H and 17-H signals in the ^1H NMR spectra of compounds **XIII** and **XV**; as a result, these signals appear in the region free from other signals, and they show a clearly defined fine structure. The 8-H signal is a doublet of triplets ($J_{8,7\beta} = 14.0$, $J_{8,9} = 4.0$ Hz) at δ 3.03 (**XIII**) and 3.04 ppm (**XV**), and the 17-H signal is a doublet of doublets ($J = 14.5$, 11.0 Hz) at δ 2.48 ppm.

EXPERIMENTAL

The IR spectra were recorded on a Specord 75IR spectrometer from samples dispersed in mineral oil. The ^1H and ^{13}C NMR spectra were measured on a Bruker AM-300 instrument (300.13 for ^1H and 75.46 MHz for ^{13}C) using CDCl_3 and CD_3OD as solvents. Homo- and heteronuclear COSY, TOCSY, ROESY, HSQC, and HBMBC experiments were run on a Bruker DRX-500 spectrometer (500.13 MHz for ^1H and 125.76 MHz for ^{13}C) using DMSO as solvent; to identify hydroxy groups, samples were subjected to lyophilization. The chemical shifts were measured relative to tetramethylsilane as internal reference. The melting points were determined on a Boetius melting point apparatus. The specific rotations were measured on a Perkin-Elmer 141 polarimeter. Thin-layer chromatography was performed on silica gel (Silufol plates); spots were visualized by treatment with a solution of vanillin in ethanol acidified with sulfuric acid.

(20R,22R)-2 β ,3 β ,14 α ,20,22,25-Hexahydroxy-7,8 α -dihydro-5 β -cholestan-6-one (IV, 7,8 α -dihydro-20-hydroxyecdysone). A suspension of 0.5 g (1.04 mmol) of 20-hydroxyecdysone **I** (mp 246°C; prepared according to the procedure described in [14]) in 20 ml of diethyl ether was added under argon to a suspension of 0.277 g (7.29 mmol) of LiAlH_4 in 30 ml of diethyl ether, cooled to $\sim 0^\circ\text{C}$. The mixture was stirred for 30 min at room temperature, 10 ml of methanol was added, the mixture was stirred for 2 h and cooled to 0°C , 5 ml of water was added, and the mixture was neutralized with 5% hydrochloric acid (~ 13 ml) to a weakly acidic reaction. The mixture was evaporated under reduced pressure to a volume of ~ 20 ml and extracted with ethyl acetate (3 \times 50 ml). The extract was evaporated under reduced pressure, and the residue was subjected to chromatography on 15 g of silica gel using chloroform-methanol (10:1) as

eluent. Yield 0.103 g (21%), R_f 0.52 (CHCl_3 -MeOH, 5:1), mp 138–140°C, $[\alpha]_{\text{D}}^{17} = +24.5^\circ$ ($c = 3.47$, MeOH) (cf. [5]). The IR and ^1H and ^{13}C NMR spectra of the product were identical to those given in [5].

(20R,22R)-14 α ,25-Dihydroxy-2 β ,3 β :20,22-bis(isopropylidenedioxy)-5 β ,8 α -cholestan-6-one (V, 7,8 α -dihydro-20-hydroxyecdysone 2,3:20,22-diacetonide). A solution of 0.82 g (1.46 mmol) of diacetonide **II** (mp 234°C; prepared as described in [15]) in 25 ml of diethyl ether was added under stirring in an argon atmosphere to a suspension of 0.16 g (4.39 mmol) of LiAlH_4 in 25 ml of diethyl ether, cooled to $\sim 0^\circ\text{C}$. The mixture was stirred for 1 h at room temperature, cooled to 0°C , and 3 ml of water and ~ 5 ml of 5% hydrochloric acid (to a weakly acidic reaction) were added. The ether layer was separated, and the aqueous layer was extracted with ethyl acetate (3 \times 30 ml). The extracts were combined with the organic phase and evaporated under reduced pressure, and the residue was subjected to chromatography on 5 g of silica gel using chloroform as eluent to isolate 0.29 g (35%) of compound **V**, R_f 0.63 (CHCl_3 -MeOH, 10:1), and 0.49 g (60%) of previously described [8] alcohols **VII** {In the ^{13}C NMR spectrum given in [8], signals from C^{24} , δ_{C} 39.6 and 39.2 ppm, respectively, for the 6 α - and 6 β -epimers of **VII** were missing}.

Compound **V**. mp 275–277°C, $[\alpha]_{\text{D}}^{19} = +14.2^\circ$ ($c = 11.97$, CHCl_3). IR spectrum: $\nu(\text{C}=\text{O})$ 1700 cm^{-1} . ^1H NMR spectrum, δ , ppm: in CDCl_3 : 1.07 s (3H, C^{18}H_3), 1.10 s (3H, C^{21}H_3), 1.19 s (6H, C^{26}H_3 , C^{27}H_3), 1.35 s (3H, C^{19}H_3), 1.26 s, 1.27 s, 1.38 s, 1.47 s (12H, Me_2C), 1.54–2.09 m (17H, CH, CH_2), 2.16 m (1H, 7 α -H), 2.29 m (1H, 17-H), 2.33 d.t (1H, 8-H, $J_{8,7\beta} = 13.5$, $J_{8,7\alpha} = J_{8,9} = 4.0$ Hz), 2.55 br.s (1H, 5-H, $w_{1/2} = 8.9$ Hz), 2.68 t (1H, 7 β -H, $J_{7\beta,7\alpha} = J_{7\beta,8} = 13.4$ Hz), 3.62 m (1H, 22-H, $w_{1/2} = 13.0$ Hz), 4.17 m (1H, 2-H, $w_{1/2} = 12$ Hz), 4.45 m (1H, 3-H, $w_{1/2} = 21$ Hz); in $\text{DMSO}-d_6$ (500.13 MHz): 1.02 s (3H, C^{18}H_3), 1.05 s (3H, C^{21}H_3), 1.07 s (3H, C^{27}H_3), 1.08 s (3H, C^{26}H_3), 1.16 m and 1.76 m (2H, 15-H), 1.19 s and 1.39 s (6H, 2,3- Me_2C), 1.23 s and 1.32 s (6H, 20,22- Me_2C), 1.28 s (3H, C^{19}H_3), 1.34 m and 1.55 m (2H, 24-H), 1.36 m and 1.39 m (2H, 23-H), 1.38 s and 1.99 s (2H, 4-H), 1.52 m and 1.67 m (2H, 1-H), 1.62 m and 1.81 m (2H, 12-H), 1.64 m and 1.77 m (2H, 11-H), 1.69 m and 1.81 m (2H, 16-H), 1.98 d.t (1H, 9-H, $J = 11.0$, 4.0 Hz), 2.06 d.d (1H, 7 α -H, $J_{7\alpha,8} = 4.0$, $J_{7\alpha,7\beta} = 13.0$ Hz), 2.20 m (1H, 17-H), 2.68 br.s (1H, 5-H, $w_{1/2} = 2$ Hz), 2.71 t (1H, 7 β -H, $J_{7\beta,7\alpha} = J_{7\beta,8} = 13.0$ Hz), 3.30 d.t (1H, 8-H, $J_{8,7\beta} = 13.0$, $J_{8,7\alpha} = J_{8,9} = 4.0$ Hz),

3.57 m (1H, 22-H), 3.92 s (1H, 14-OH), 4.13 s (1H, 25-OH), 4.14 m (1H, 2-H), 4.27 m (1H, 3-H). ^{13}C NMR spectrum, δ_{C} , ppm: in CDCl_3 : 18.1 t (C^{11}), 18.4 q (C^{18}), 21.0 t (C^{16}), 21.3 q (C^{21}), 23.5 t (C^{23}), 25.3 t (C^4), 26.7 q (C^{19}), 25.8 q and 28.4 q (2,3- Me_2C), 26.7 q and 28.9 q (20,22- Me_2C), 29.0 q (C^{27}), 29.6 q (C^{26}), 31.3 t (C^{15}), 33.1 t (C^{12}), 34.2 t (C^1), 39.5 s (C^{10}), 41.3 t (C^{24}), 41.3 t (C^7), 41.5 d (C^9), 43.7 d (C^8), 46.8 s (C^{13}), 49.7 d (C^{17}), 50.5 d (C^5), 70.2 s (C^{25}), 70.8 d (C^3), 73.5 d (C^2), 81.8 d (C^{22}), 84.4 s (C^{14}), 84.9 s (C^{20}), 106.8 s (20,22- Me_2C), 107.6 s (2,3- Me_2C), 212.1 s (C^6); in DMSO (500.13 MHz): 17.8 t (C^{11}), 18.0 q (C^{18}), 21.0 t (C^{16}), 21.4 q (C^{21}), 23.1 t (C^{23}), 25.3 t (C^4), 25.9 q (C^{19}), 25.9 q and 28.5 q (2,3- Me_2C), 26.6 q and 29.0 q (20,22- Me_2C), 29.0 q (C^{27}), 29.7 q (C^{26}), 30.1 t (C^{15}), 32.8 t (C^{12}), 33.8 t (C^1), 38.9 s (C^{10}), 40.8 t (C^7), 40.9 d (C^9), 41.1 t (C^{24}), 42.7 d (C^8), 46.3 s (C^{13}), 49.3 d (C^{17}), 49.5 d (C^5), 68.4 s (C^{25}), 70.2 d (C^3), 73.0 d (C^2), 81.3 d (C^{22}), 82.8 s (C^{14}), 84.1 s (C^{20}), 105.8 s (20,22- Me_2C), 106.8 s (2,3- Me_2C), 211.9 s (C^6).

(20R,22R)-14 α -Hydroxy-2 β ,3 β :20,22-bis(isopropylidenedioxy)-5 β ,8 α -cholest-24(25)-en-6-ones [VI, 24,25(25,26)-anhydro-7,8 α -dihydro-20-hydroxyecdysone 2,3:20,22-diacetonides] (mixture of isomers). A solution of 1.3 g (2.40 mmol) of isomeric alkenes **III** (prepared as described in [12, 16]) in 30 ml of diethyl ether was added under stirring in an argon atmosphere to a suspension of 0.27 g (7.2 mmol) of LiAlH_4 in 30 ml of diethyl ether, cooled to $\sim 0^\circ\text{C}$. The mixture was stirred for 3 h at room temperature, cooled to 0°C , 5 ml of water was added, and ~ 10 ml of 5% hydrochloric acid was added to a weakly acidic reaction. The organic layer was separated, and the aqueous layer was extracted with ethyl acetate (3×50 ml). The extracts were combined with the organic phase and evaporated under reduced pressure, and the residue was subjected to chromatography on 30 g of silica gel using chloroform as eluent to isolate 0.39 g (30%) of compound **VI**, R_f 0.5 (CHCl_3 -MeOH, 20:1), and 0.85 g (65%) of previously described [8] alcohols **VIII**.

Compound (**VI**). mp 190 – 192°C , $[\alpha]_{\text{D}}^{19} = +16.1^\circ$ ($c = 8.83$, CHCl_3). IR spectrum, ν , cm^{-1} : 1695 ($\text{C}=\text{O}$). ^1H NMR spectrum (CDCl_3), δ , ppm: 0.91 s and 0.96 s (3H, C^{18}H_3 , $\text{C}^{18'}\text{H}_3$); 0.94 s (3H, C^{19}H_3); 1.11 s and 1.14 s (3H, C^{21}H_3 , $\text{C}^{21'}\text{H}_3$); 1.28 s, 1.38 s, 1.40 s, and 1.50 s (3H each, Me_2C); 1.78–2.20 m (13H, CH, CH_2); 1.62 s ($\sim 2.7\text{H}$, C^{26}H_3); 1.70 s and 1.72 s (3H, C^{27}H_3 , $\text{C}^{27'}\text{H}_3$); 2.22 m (1H, 7 α -H); 2.30 m (1H, 17-H); 2.37 m (1H, 8-H); 2.57 br.s (1H, 5-H, $w_{1/2} = 10.7$ Hz);

2.70 t (1H, 7 β -H, $J = 13.4$ Hz); 3.65–3.71 m (1H, 22-H, 22'-H); 4.20 m (1H, 2-H, $w_{1/2} = 13.0$ Hz); 4.49 m (1H, 3-H, $w_{1/2} = 22.6$ Hz); 4.69 br.s and 4.72 br.s (1.3H, 26-H, $w_{1/2} = 6.8$ Hz); 5.17 t (0.7H, 24'-H, $J = 6.2$ Hz). ^{13}C NMR spectrum (CDCl_3), δ_{C} , ppm: 17.9 q (C^{18}), 18.2 t (C^{11}), 18.6 q ($\text{C}^{27'}$), 21.1 t (C^{16}), 21.4 q (C^{21}), 22.5 q (C^{27}), 25.4 t (C^{23}), 25.7 q (C^{19}), 25.9 q (C^{26}), 26.0 q and 28.5 q (2,3- Me_2C), 26.8 q and 29.6 q (20,22- Me_2C), 27.7 t (C^4), 31.5 t (C^{15}), 33.3 t (C^{12}), 34.4 t (C^{24}), 35.0 t (C^1), 39.6 s (C^{10}), 41.3 t (C^7), 41.6 d and 41.7 d (C^9), 43.8 d and 43.9 d (C^8), 46.9 s (C^{13}), 50.0 d and 50.1 d (C^{17}), 50.6 d (C^5), 70.9 d (C^3), 73.5 d (C^2), 80.6 d and 81.0 d (C^{22}), 84.1 s (C^{20}), 85.0 s (C^{14}), 106.7 s and 106.8 s (20,22- Me_2C), 107.6 s (2,3- Me_2C), 110.0 t (C^{26}), 120.4 d (C^{24}), 133.5 q (C^{25}), 145.3 s (C^{25}), 211.9 s (C^6).

(20R,22R)-2 β ,3 β :20,22-Bis(isopropylidenedioxy)-5 α -cholesta-7,14-diene-6 β ,25-diol (IX). Diacetone **II**, 1 g (1.79 mmol), was dissolved in 50 ml of anhydrous THF, 0.198 g (5.37 mmol) of LiAlH_4 was added, and the mixture was stirred for 1 h at $\sim 25^\circ\text{C}$ under argon. The mixture was cooled to 0°C , 5 ml of water was added, and ~ 7 ml of 5% hydrochloric acid was added to a weakly acidic reaction. The organic layer was separated, and the aqueous layer was extracted with ethyl acetate (3×70 ml). The extracts were combined with the organic phase and evaporated under reduced pressure, and the residue was subjected to chromatography on 40 g of silica gel using chloroform as eluent to isolate 0.1 g (10%) of dienol **IX**, R_f 0.7 (CHCl_3 -MeOH, 10:1), 0.3 g (30%) of **V**, R_f 0.63 (CHCl_3 -MeOH, 10:1), and 0.53 g (53%) of **VII**.

Dienol (**IX**). mp 90 – 92°C , $[\alpha]_{\text{D}}^{19} = -62.8^\circ$ ($c = 7.32$, CHCl_3). ^1H NMR spectrum (CDCl_3), δ , ppm: 1.01 s (3H, C^{18}H_3); 1.04 s (3H, C^{19}H_3); 1.18 s (3H, C^{21}H_3); 1.22 s (6H, C^{26}H_3 , C^{27}H_3); 1.29 s, 1.31 s, 1.41 s, and 1.50 s (3H each, Me_2C); 1.52–2.58 m (15H, CH, CH_2); 3.72 m (1H, 22-H, $w_{1/2} = 15.3$ Hz); 3.81 m (1H, 6-H, $w_{1/2} = 10$ Hz); 4.05 m (1H, 2-H, $w_{1/2} = 20$ Hz); 4.19 m (1H, 3-H, $w_{1/2} = 11$ Hz); 5.62 br.s (1H, 15-H, $w_{1/2} = 8$ Hz); 5.89 br.s (1H, 7-H, $w_{1/2} = 9$ Hz). ^{13}C NMR spectrum (CDCl_3), δ_{C} , ppm: 18.4 q (C^{18}), 20.9 t (C^{11}), 21.1 q (C^{21}), 23.7 t (C^{23}), 24.8 q (C^{19}), 28.3 t (C^4), 29.0 q (C^{27}), 29.6 q (C^{26}), 26.5 q and 28.6 q (2,3- Me_2C), 26.8 q and 28.8 q (20,22- Me_2C), 30.9 t (C^{12}), 34.3 s (C^{10}), 37.2 d (C^9), 39.2 t (C^{16}), 40.4 t (C^{24}), 41.2 t (C^1), 43.3 d (C^5), 47.3 s (C^{13}), 57.6 d (C^{17}), 70.3 s (C^{25}), 70.8 d (C^6), 72.3 d (C^2), 72.8 d (C^3), 81.7 d (C^{22}), 83.6 s (C^{20}), 106.8 s (20,22- Me_2C), 107.9 s (2,3- Me_2C), 120.3 d (C^{15}), 121.2 d (C^7), 135.2 s (C^{14}), 150.0 s (C^8).

(20R,22R)-2 β ,3 β :20,22-Bis(isopropylidenedioxy)-5 β ,8 α -cholest-14-ene-6 β ,25-diol (X). Diacetonide **II**, 0.3 g (0.54 mmol), was dissolved in 20 ml of anhydrous THF, 0.1 g (2.70 mmol) of LiAlH₄ was added, and the mixture was stirred for 1 h at ~25°C under argon. The mixture was cooled to 0°C, 5 ml of water was added, and ~5 ml of 5% hydrochloric acid was added to a weakly acidic reaction. The organic layer was separated, and the aqueous phase was extracted with ethyl acetate (3×50 ml). The extracts were combined with the organic phase and evaporated under reduced pressure, and the residue was subjected to chromatography on 20 g of silica gel using chloroform as eluent to isolate 0.05 g (17%) of diol **X**, *R_f* 0.69 (CHCl₃-MeOH, 10:1), 0.088 g (30%) of **V**, *R_f* 0.63 (CHCl₃-MeOH, 10:1), and 0.15 g (50%) of **VII**.

Diol (**X**). mp 75–77°C, $[\alpha]_D^{24} = +35.6^\circ$ (*c* = 2.74, CHCl₃). ¹H NMR spectrum (CDCl₃), δ , ppm: 1.13 s (3H, C¹⁸H₃); 1.16 s (3H, C¹⁹H₃); 1.18 s (3H, C²¹H₃); 1.22 s (6H, C²⁶H₃, C²⁷H₃); 1.30 s, 1.41 s, and 1.51 s (2:1:1, 12H, Me₂C); 1.60–2.52 m (20H, CH, CH₂); 3.07 m (1H, 8-H); 3.75 m (1H, 22-H, *w*_{1/2} = 21 Hz); 3.95 br.s (1H, 6-H, *w*_{1/2} = 12 Hz); 4.36 m (1H, 2-H, *w*_{1/2} = 13 Hz); 4.63 m (1H, 3-H, *w*_{1/2} = 16 Hz); 5.35 br.s (1H, 15-H, *w*_{1/2} = 13 Hz). ¹³C NMR spectrum (CDCl₃), δ_C , ppm: 17.7 t (C¹¹), 20.8 q (C¹⁸), 23.7 t (C²³), 25.8 q (C²¹), 26.7 q and 28.9 q (2,3-Me₂C), 27.3 q and 28.9 q (20,22-Me₂C), 28.6 q (C¹⁹), 29.7 q (C²⁶, C²⁷), 30.5 t (C¹⁶), 31.0 d (C⁹), 31.6 t (C⁴), 33.4 t (C¹⁰), 36.0 t (C⁷), 37.9 t (C¹), 41.0 d (C⁵), 41.4 s (C¹²), 43.1 t (C²⁴), 46.8 s (C¹³), 48.2 d (C⁸), 59.5 d (C¹⁷), 70.3 s (C²⁵), 71.6 d (C⁶), 72.7 d (C³), 74.6 d (C²), 81.8 d (C²²), 83.7 s (C²⁰), 106.7 s (20,22-Me₂C), 107.0 s (2,3-Me₂C), 120.6 d (C¹⁵), 154.4 s (C¹⁴).

(20R,22R)-2 β ,3 β :20,22-Bis(isopropylidenedioxy)-5 β ,8 α -cholestane-6 α ,14 α ,25-triol (XI). Diacetonide **V**, 0.18 g (0.32 mmol), was dissolved in 7 ml of anhydrous THF, 0.025 g (0.64 mmol) of LiAlH₄ was added, and the mixture was stirred for 0.5 h at ~25°C under argon. The mixture was cooled to 0°C, 5 ml of water was added, and ~7 ml of 5% hydrochloric acid was added to a weakly acidic reaction. The organic layer was separated, and the aqueous layer was extracted with ethyl acetate (3×40 ml). The extracts were combined with the organic phase and evaporated under reduced pressure, and the residue was subjected to chromatography on 8 g of silica gel using chloroform as eluent. Yield 0.16 g (90%), *R_f* 0.34 (CHCl₃-MeOH, 10:1), mp 137–139°C, $[\alpha]_D^{20} = +25.7^\circ$ (*c* = 9.33, CHCl₃). ¹H NMR spectrum, δ , ppm: in CDCl₃: 1.00 s

(3H, C¹⁸H₃); 1.08 s (3H, C²¹H₃); 1.18 s (9H, C¹⁹H₃, C²⁶H₃, C²⁷H₃); 1.27 s, 1.37 s, and 1.47 s (2:1:1, 12H, Me₂C); 1.60–2.00 m (20H, CH, CH₂); 2.13 m (1H, 17-H, *w*_{1/2} = 19 Hz); 2.32 m (1H, 8-H, *w*_{1/2} = 24 Hz); 3.62 m (1H, 22-H, *w*_{1/2} = 16 Hz); 3.94 br.s (1H, 6-H, *w*_{1/2} = 10 Hz); 4.29 m (1H, 2-H, *w*_{1/2} = 12 Hz); 4.58 m (1H, 3-H, *w*_{1/2} = 16 Hz); in DMSO-*d*₆ (500.13 MHz): 0.92 s (3H, C¹⁸H₃), 1.01 s (3H, C²¹H₃), 1.06 s (C²⁷H₃), 1.07 s (6H, C¹⁹H₃, C²⁶H₃), 1.18 s and 1.37 s (6H, 2,3-Me₂C), 1.22 s and 1.31 s (6H, 20,22-Me₂C), 1.15 m and 1.79 m (2H, 15-H), 1.30 m and 2.33 m (2H, 1-H), 1.34 m and 1.37 m (2H, 23-H), 1.31 m and 1.54 m (2H, 24-H), 1.35 m and 1.39 m (2H, 11-H), 1.51 m and 1.71 m (2H, 12-H), 1.46 m and 1.71 m (2H, 7-H), 1.63 m and 1.68 m (2H, 4-H), 1.37 m (1H, 5-H), 1.66 m and 1.79 m (2H, 16-H), 1.81 m (1H, 9-H), 2.20 m (1H, 17-H), 2.31 m (1H, 8-H), 3.56 m (1H, 22-H), 3.65 s (1H, 14-OH), 3.75 m (1H, 6-H), 4.13 s (1H, 25-OH), 4.19 m (1H, 2-H), 4.39 d (1H, 6-OH, *J* = 3.5 Hz), 4.51 m (1H, 3-H). ¹³C NMR spectrum, δ_C , ppm: in CDCl₃: 18.0 t (C¹¹); 18.3 q (C¹⁸); 21.4 t (C¹⁶); 23.5 t (C²³); 25.9 q (C²¹); 26.7 q (C¹⁹); 27.6 q, 28.6 q, 28.8 q, and 28.9 q (Me₂C); 29.6 q (C²⁶, C²⁷); 31.2 t (C¹⁵); 31.5 t (C⁴); 33.4 s (C⁷, C¹⁰); 34.0 t (C¹²); 36.5 t (C¹); 36.7 d (C⁸); 41.3 t (C²⁴); 42.3 d (C⁵, C⁹); 46.6 s (C¹³); 49.8 d (C¹⁷); 70.2 s (C²⁵); 71.8 d (C⁶); 72.6 d (C³); 74.6 s (C²); 81.8 d (C²²); 84.6 s (C¹⁴); 86.1 s (C²⁰); 106.7 s (20,22-Me₂C); 107.0 s (2,3-Me₂C); in DMSO (500.13 MHz): 17.9 t (C¹¹), 18.1 q (C¹⁸), 21.2 t (C¹⁶), 21.4 q (C²¹), 23.2 t (C²³), 25.9 q and 28.6 q (2,3-Me₂C), 26.7 q and 29.0 q (20,22-Me₂C), 27.3 q (C¹⁹), 29.0 q (C²⁷), 29.6 q (C²⁶), 30.4 t (C¹⁵), 31.7 t (C⁴), 32.8 s (C⁷), 33.0 s (C¹⁰), 33.6 t (C¹²), 35.9 d (C⁸), 36.2 t (C¹), 41.1 t (C²⁴), 41.2 d (C⁵), 41.5 d (C⁹), 46.1 s (C¹³), 49.3 d (C¹⁷), 68.4 s (C²⁵), 70.3 d (C⁶), 71.9 d (C³), 74.2 s (C²), 81.3 d (C²²), 83.9 s (C¹⁴), 84.3 s (C²⁰), 105.8 s (20,22-Me₂C), 106.0 s (2,3-Me₂C).

(20R,22R)-2 β ,3 β ,14 α ,25-Tetrahydroxy-20,22-isopropylidenedioxy-5 β ,8 α -cholestan-6-one (XII, 7,8 α -dihydro-20-hydroxyecdysone 20,22-acetonide (XII)). A mixture of 0.22 g (0.46 mmol) of compound **V** and 3 ml of glacial acetic acid was stirred for 5.5 h. The mixture was evaporated, and the residue was subjected to chromatography on 9 g of silica gel using chloroform-methanol (20:1) as eluent. Yield 0.1 g (49%), *R_f* 0.60 (CHCl₃-MeOH, 5:1), mp 134–136°C, $[\alpha]_D^{18} = +7.2^\circ$ (*c* = 8.67, CHCl₃). IR spectrum: ν (C=O) 1700 cm⁻¹. ¹H NMR spectrum (CDCl₃), δ , ppm: 1.06 s (3H, C¹⁸H₃), 1.11 s (3H, C²¹H₃), 1.21 s (6H, C²⁶H₃, C²⁷H₃), 1.29 s (3H, C¹⁹H₃), 1.39 s (6H, Me₂C), 1.42–2.10 m (19H, CH, CH₂), 2.31 t (1H, 8-H, *J* = 12.5 Hz), 2.43 m

(1H, 5-H, $w_{1/2} = 11$ Hz), 2.67 t (1H, 7 β -H, $J = 14.5$ Hz), 3.63 m (1H, 22-H, $w_{1/2} = 15$ Hz), 3.84 m (1H, 2-H, $w_{1/2} = 18$ Hz), 3.93 m (1H, 3-H, $w_{1/2} = 12$ Hz). ^{13}C NMR spectrum (CDCl_3), δ_{C} , ppm: 18.4 q (C^{18}), 20.1 t (C^{11}), 21.1 t (C^{16}), 21.4 q (C^{21}), 23.4 t (C^{23}), 25.0 t (C^4), 26.7 q (C^{19}), 26.9 q and 28.9 q (20,22- Me_2C), 29.1 q (C^{27}), 29.4 q (C^{26}), 31.3 t (C^{15}), 33.5 t (C^{12} , C^1), 39.4 s (C^{10}), 41.0 d (C^9), 41.1 t (C^7), 41.3 t (C^{24}), 43.0 d (C^8), 46.8 s (C^{13}), 49.8 d (C^{17}), 51.1 d (C^5), 70.4 s (C^{25}), 66.7 d (C^3), 69.5 d (C^2), 81.9 d (C^{22}), 84.4 s (C^{20}), 85.0 s (C^{14}), 106.8 s (20,22- Me_2C), 212.8 s (C^6).

(20R,22R)-2 β ,3 β ,25-Trihydroxy-20,22-isopropylidenedioxy-5 β ,8 α -cholest-14-en-6-one (XIII, 7,8 α -dihydrostachysterone B 20,22-acetonide). A mixture of 0.125 g (0.22 mmol) of compound V and 3 ml of 70% acetic acid was stirred for 1.5 h, 0.045 g (0.33 mmol) of ZnCl_2 was added, and the mixture was stirred for 2.5 h until the reaction was complete (TLC). The mixture was diluted with 30 ml of water and extracted with butanol (3 \times 20 ml), the extracts were combined and evaporated under reduced pressure, and the residue was subjected to chromatography on 8 g of silica gel using chloroform–methanol (20:1) as eluent. Yield 0.067 g (60%), R_f 0.47 (CHCl_3 –MeOH, 20:1), mp 118–120°C, $[\alpha]_{\text{D}}^{24} = -123^\circ$ ($c = 1.18$, CHCl_3). IR spectrum: $\nu(\text{C}=\text{O})$ 1700 cm^{-1} . ^1H NMR spectrum (CDCl_3), δ , ppm: 1.16 s (6H, C^{18}H_3 , C^{21}H_3), 1.21 s (6H, C^{26}H_3 , C^{27}H_3), 1.28 s (3H, C^{19}H_3), 1.40 s and 1.41 s (3H each, Me_2C), 1.46–2.21 m (15H, CH, CH_2), 2.15 d.d (1H, 7 α -H, $J = 14.5$, 4.4 Hz), 2.36 m (1H, 5-H, $w_{1/2} = 10$ Hz), 2.48 d.d (1H, 17-H, $J = 14.5$, 11.0 Hz), 2.73 t (1H, 7 β -H, $J = 14.5$ Hz), 3.03 d.t (1H, 8-H, $J = 14.5$, 4.0 Hz), 3.72 m (1H, 22-H, $w_{1/2} = 5$ Hz), 3.87 m (1H, 2-H, $w_{1/2} = 24$ Hz), 3.97 m (1H, 3-H, $w_{1/2} = 11$ Hz), 5.33 m (1H, 15-H, $w_{1/2} = 8$ Hz). ^{13}C NMR spectrum (CDCl_3), δ_{C} , ppm: 17.9 t (C^{11}), 21.0 q (C^{18} , C^{21}), 23.6 t (C^{23}), 24.3 t (C^{16}), 26.7 q (C^{19}), 28.8 q (20,22- Me_2C), 29.0 q (C^{26}), 29.5 q (C^{27}), 30.4 t (C^4), 37.3 d (C^9), 38.4 t (C^1), 39.8 s (C^{10}), 41.2 t (C^{24}), 42.4 t (C^{12}), 44.2 t (C^7), 46.7 s (C^{13}), 47.3 d (C^8), 51.2 d (C^5), 59.3 d (C^{17}), 70.3 s (C^{25}), 66.7 d (C^3), 69.5 d (C^2), 81.6 d (C^{22}), 83.4 s (C^{20}), 106.8 s (20,22- Me_2C), 122.2 d (C^{15}), 151.1 s (C^{14}), 211.9 s (C^6).

(20R,22R)-2 β ,3 β ,20,22,25-Pentahydroxy-5 β ,8 α -cholest-14-en-6-one (XIV, 7,8 α -dihydrostachysterone B). Compound V, 0.2 g (0.36 mmol), was dissolved in 3.6 ml of methanol, 1 ml of 10% perchloric acid was added, the mixture was stirred for 3.5 h and cooled to 5°C, 2 ml of water and 1 ml of a saturated

solution of NaHCO_3 were added, and the mixture was extracted with ethyl acetate (3 \times 20 ml). The extracts were combined and evaporated under reduced pressure, and the residue was subjected to chromatography on 8 g of silica gel using chloroform–methanol (20:1) as eluent. Yield 0.07 g (33%), R_f 0.38 (CHCl_3 –MeOH, 5:1), mp 112–114°C, $[\alpha]_{\text{D}}^{18} = -56.3^\circ$ ($c = 1.1$, MeOH). IR spectrum: $\nu(\text{C}=\text{O})$ 1700 cm^{-1} . ^1H NMR spectrum (CD_3OD), δ , ppm: 1.17 s (3H, C^{18}H_3), 1.19 s (3H, C^{19}H_3), 1.20 s (3H, C^{21}H_3), 1.26 s (3H, C^{26}H_3), 1.42 s (3H, C^{27}H_3), 1.50–2.40 m (17H, CH, CH_2), 2.48 m (1H, 5-H, $w_{1/2} = 11$ Hz), 2.88 t (1H, 7 β -H, $J = 14$ Hz), 3.07 m (1H, 8-H, $w_{1/2} = 25$ Hz), 3.30 m (1H, 22-H), 3.74 m (1H, 2-H, $w_{1/2} = 19$ Hz), 3.92 m (1H, 3-H, $w_{1/2} = 10$ Hz), 5.39 br.s (1H, 15-H, $w_{1/2} = 8$ Hz). ^{13}C NMR spectrum (CD_3OD), δ_{C} , ppm: 19.2 t (C^{11}), 20.1 q (C^{21}), 21.9 q (C^{18}), 25.3 t (C^{23}), 27.3 t (C^{16}), 27.5 q (C^{19}), 28.9 q (C^{26}), 29.4 q (C^{27}), 30.7 t (C^4), 38.9 d (C^9), 40.3 t (C^1), 41.0 s (C^{10}), 42.3 t (C^{12} , S^{24}), 44.5 t (C^7), 45.4 d (C^8), 52.4 d (C^5), 60.8 d (C^{17}), 71.3 s (C^{25}), 68.1 d (C^3), 71.0 d (C^2), 77.2 s (C^{20}), 78.6 d (C^{22}), 123.7 d (C^{15}), 152.9 s (C^{14}), 214.0 s (C^6); the C^{13} signal was obscured by the solvent.

(20R,22R)-2 β ,3 β -Dihydroxy-20,22-isopropylidenedioxy-5 β ,8 α -cholest-14-en-6-one (XV). Hydrogen was passed through a suspension of 0.26 g (0.48 mmol) of compound VI and 0.1 g (10%) of Pd/C in 5 ml of ethanol under stirring at $\sim 25^\circ\text{C}$. After 3 days, the mixture was filtered, the catalyst was washed with ethanol, and the filtrate was evaporated under reduced pressure. The residue was subjected to chromatography on 6 g of silica gel using chloroform as eluent. Yield 0.094 g (41%), R_f 0.39 (CHCl_3 –MeOH, 20:1), mp 110–112°C, $[\alpha]_{\text{D}}^{23} = -18.6^\circ$ ($c = 11.15$, CHCl_3). IR spectrum: $\nu(\text{C}=\text{O})$ 1710 cm^{-1} . ^1H NMR spectrum (CDCl_3), δ , ppm: 0.87 d (6H, C^{26}H_3 , C^{27}H_3 , $J = 6.5$ Hz), 1.14 s (3H, C^{21}H_3), 1.17 s (3H, C^{18}H_3), 1.27 s (3H, C^{19}H_3), 1.40 s and 1.42 s (6H, Me_2C), 1.46–2.22 m (16H, CH, CH_2), 2.15 d.d (1H, 7 α -H, $J = 14.5$, 3.4 Hz), 2.37 m (1H, 9-H, $w_{1/2} = 11$ Hz), 2.49 d.d (1H, 5-H, $J = 11.7$, 14.8 Hz), 2.74 t (1H, 7 β -H, $J = 14.5$ Hz), 3.04 d.t (1H, 8-H, $J = 13.8$, 5.1 Hz), 3.69 d.d (1H, 22-H, $J = 8.6$, 2.5 Hz), 3.91 m (1H, 3-H, $w_{1/2} = 24$ Hz), 3.99 m (1H, 2-H, $w_{1/2} = 11$ Hz), 5.35 m (1H, 15-H, $w_{1/2} = 6$ Hz). ^{13}C NMR spectrum (CDCl_3), δ_{C} , ppm: 17.9 t (C^{11}), 21.0 q (C^{18}), 21.0 q (C^{21}), 22.4 q (C^{26}), 22.4 q (C^{27}), 24.3 t (C^{23}), 26.7 q (C^{19}), 26.8 t (C^{16}), 28.9 s (C^{25}), 28.1 q and 28.9 q (20,22- Me_2C), 30.4 t (C^4), 36.2 t (C^{24}), 37.4 d (C^9), 38.3 t (C^1), 39.8 s (C^{10}), 42.5 t (C^{12}), 44.2 t (C^7), 46.8 s (C^{13}), 47.3 d (C^8),

51.2 d (C⁵), 59.5 d (C¹⁷), 69.6 d (C³), 70.7 d (C²), 81.3 d (C²²), 83.2 s (C²⁰), 106.6 s (20,22-Me₂C), 122.3 d (C¹⁵), 151.1 s (C¹⁴), 212.0 s (C⁶).

This study was performed under financial support by the Russian Foundation for Basic Research (project no. 04-03-33103) and by the President of the Russian Federation (program for state support of young Russian scientists, project no. MK-6975.2006.3).

REFERENCES

1. Afon'kina, S.R., Shafikov, R.V., Savchenko, R.G., Galyautdinov, I.V., and Odinokov, V.N., *Russ. J. Org. Chem.*, 2006, vol. 42, p. 1234.
2. Akhrem, A.A. and Kovganko, N.V., *Ekdisteroidy: khimiya i biologicheskaya aktivnost'* (Ecdysteroids: Chemistry and Biological Activity), Minsk: Nauka i Tekhnika, 1989.
3. Werawattanametin, K., Podimnang, V., and Suksamrarn, A., *J. Nat. Prod.*, 1986, vol. 49, p. 365.
4. Odinokov, V.N., Galyautdinov, I.V., Nedopekin, D.V., Ves'kina, N.A., and Khalilov, L.M., *Russ. J. Org. Chem.*, 2003, vol. 39, p. 952.
5. Suksamrarn, A., Tanachatchairatana, T., and Sirigarn, C., *Tetrahedron*, 2002, vol. 58, p. 6033.
6. Caine, D., *Organic Reactions*, Dauben, W.G., Ed., New York: Wiley, 1976, vol. 23.
7. Dryden, H.L., Jr., *Organic Reactions in Steroid Chemistry*, Fried, J. and Edwards, J.A., New York: Van Nostrand Reinhold, 1972, vol. 1, p. 60.
8. Odinokov, V.N., Savchenko, R.G., Shafikov, R.V., Afon'kina, S.R., Khalilov, L.M., Kachala, V.V., and Shashkov, A.S., *Russ. J. Org. Chem.*, 2005, vol. 41, p. 1296.
9. Greenwood, D.R., Dinan, L.N., and Rees, H.H., *Biochem. J.*, 1984, vol. 217, p. 783.
10. Girault, J.-P., Blais, C., Beydon, P., Rolando, C., and Lafont, R., *Arch. Insect Biochem. Physiol.*, 1989, vol. 10, p. 199.
11. Ionin, B.I., Ershov, B.A., and Kol'tsov, A.I., *YaMR-spektroskopiya v organicheskoi khimii* (NMR Spectroscopy in Organic Chemistry), Leningrad: Khimiya, 1983.
12. Yingyongnarongkul, B. and Suksamrarn, A., *Tetrahedron*, 1998, vol. 54, p. 2795.
13. Lee, Sh.-Sh., Nakanishi, K., and Cherbas, P., *J. Chem. Soc., Chem. Commun.*, 1991, p. 51.
14. Odinokov, V.N., Galyautdinov, I.V., Nedopekin, D.V., Khalilov, L.M., Shashkov, A.S., Kachala, V.V., Dinan, L., and Lafont, R., *Insect Biochem. Molec. Biol.*, 2002, vol. 32, p. 161.
15. Odinokov, V.N., Galyautdinov, I.V., Nedopekin, D.V., and Khalilov, L.M., *Izv. Ross. Akad. Nauk, Ser. Khim.*, 2003, p. 220.
16. Odinokov, V.N., Savchenko, R.G., Nazmeeva, S.R., Galyautdinov, I.V., and Khalilov, L.M., *Russ. J. Org. Chem.*, 2002, vol. 38, p. 525.